



European Union Reference Laboratory  
for halogenated POPs in Feed and Food



State Institute for Chemical and Veterinary Analysis of Food, Freiburg, Germany

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**EURL Proficiency Test on the Determination of  
PCDD/Fs, PCBs, BFRs, PFASs and CPs  
in Fish fillet  
2020**

*EURL-PT-POP-2001-FI*

**FOOD**

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**Report  
PCDD/Fs and PCBs  
(Version 1.0)**

**24 September 2020**



This report on the EURL Proficiency Test on the Determination of PCDD/Fs, PCBs, BFRs, PFASs and CPs in Fish fillet 2020 [EURL-PT-POP\_2001-FI] organized by the EURL for halogenated Persistent Organic Pollutants (POPs) in Feed and Food is only available as pdf-version. The forwarding and reproduction of this report is permitted only as entire document, including 8 annexes.

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## Summary

<b>Test samples (food)</b>	Fish fillet (fresh water fish) - 2001-FI
<b>Analytes of interest</b>	<u>Mandatory for NRLs:</u> - PCDD/Fs (17 2,3,7,8-substituted PCDD/Fs) - PCBs (12 DL-PCBs, 6 NDL-PCBs)
<b>Methods</b>	<u>PCDD/Fs, DL-PCBs:</u> GC-HRMS, GC-MS/MS and alternative methods; Bioanalytical screening methods <u>Indicator PCBs:</u> Any kind of method
<b>Participants</b>	NRLs, OFLs, other official laboratories, commercial laboratories performing the analysis of samples taken by food business operators
<b>Statistical evaluation</b>	ISO 13528:2015, IUPAC Protocol, Positive scoring system
<b>Report</b>	24 September 2020



## 1. Structure of the PT, test material and analytes

This proficiency test (PT) on the determination of PCDD/Fs, PCBs, BFRs, PFASs and CPs in fish fillet (fresh water fish) was organized by the EURL for Halogenated Persistent Organic Pollutants (POPs) in Feed and Food to be performed between February and September 2020. The objective was to assess analytical performance of laboratories and the interlaboratory comparability of results from analyses of PCDD/Fs, PCBs, BFRs, PFASs and CPs in one sample of fish fillet.

National Reference Laboratories (NRLs) for Halogenated POPs in Feed and Food from EU member states were requested to participate as part of their work programme for 2020. NRLs were invited to encourage the participation of Official Laboratories (OFLs) from their member states as part of their duties following Article 101 of regulation (EU) 2017/625 of the European Parliament and of the Council of 15 March 2017. Furthermore, participation of OFLs will allow the extension of the data basis for calculation of assigned values and evaluation of results.

This PT was also open for other official laboratories and commercial laboratories performing the analysis of samples taken by food business operators in order to check the comparability of results not only within the EURL/NRL/OFL network, but also with official and private laboratories performing official control or self-control of food business operators.

The evaluated results will be discussed by representatives of EU Commission, NRLs and the EURL at the COM/EURL/NRL workshop in November 2020.

### 1.1 Samples and coding

The fish fillet test sample was prepared of regular market food. The test sample was not fortified with analytes of interest.

Fish fillet (fresh water fish)	Sample no. 2001-FI-xxx
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Each participant received about 125 g of the test sample.



## 1.2 Analytes of interest

NRLs for halogenated POPs in feed and food are requested to determine the following parameters:

### PCDD/Fs and PCBs:

- 17 2,3,7,8-substituted PCDD/Fs
- WHO-PCDD/F-TEQ (using WHO<sub>2005</sub>-TEF)
- 12 dioxin-like PCBs
- WHO-PCB-TEQ (using WHO<sub>2005</sub>-TEF)
- WHO-PCDD/F-PCB-TEQ (using WHO<sub>2005</sub>-TEF)
- Six indicator PCBs: PCB 28, 52, 101, 138, 153, 180
- Sum of six indicator PCBs: Sum of PCB 28, 52, 101, 138, 153, 180
- PCDD/F-PCB-BEQ, PCDD/F-BEQ and/or PCB-BEQ using bioanalytical screening methods, if applicable

## 1.3 Coding of laboratories and confidentiality

The laboratory code of the participating laboratories will be kept confidential and will not be revealed to other participants.

For NRLs, the “Protocol for management of underperformance in comparative testing and/or lack of collaboration of National Reference Laboratories (NRLs) with Community reference laboratories (CRLs) activities” will be observed. The confidentiality of NRLs will be kept according to this protocol.

The identity of OFLs will be kept confidential, unless a Member State initiated a co-operation between the NRL, OFLs and the EURL.

## 1.4 Results of PCDD/Fs and PCBs determined by physico-chemical methods

Laboratories should:

- use their own reference standards for identification and quantification,
- report results for each analyte,
- report the limit of quantification (LOQ), at least for each non-quantified analyte,
- report upper, middle and lower bound results for WHO-PCDD/F-PCB-TEQ, WHO-PCDD/F-TEQ, WHO-PCB-TEQ and sum of six indicator PCBs,
- report if sample exceeds respective EU maximum or action levels for WHO-PCDD/F-PCB-TEQ, WHO-PCDD/F-TEQ and/or WHO-PCB-TEQ or the maximum



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level for the sum of six indicator PCBs beyond reasonable doubt taking into account the measurement uncertainty,

- report the measurement uncertainty, applied for checking of compliance, for WHO-PCDD/F-PCB-TEQ, WHO-PCDD/F-TEQ, WHO-PCB-TEQ and the sum of six indicator PCBs,
- give method information and
- give information about the accreditation of the laboratory according to ISO/IEC 17025 (*for metrological traceability of consensus values of participants used as assigned values*).

Results had to be reported in pg/g wet weight for PCDD/Fs and dioxin-like PCBs, and in ng/g wet weight for indicator PCBs. TEQ-based results had to be calculated using the WHO-TEFs of 2005 [3].

## 1.5 Results of PCDD/Fs and PCBs determined by bioanalytical screening methods

Laboratories should:

- use their own reference standards;
- report if the samples are suspected to be noncompliant with EU legal limits and confirmation is required
- report PCDD/F and/or PCB results in BEQ, if applicable,
- report the reporting limit, maximum level / action level, which the evaluation is based on, and the bioassay cut-off, if applicable,
- give method information
- give information about the accreditation of the laboratory according to ISO/IEC 17025.

Results had to be reported in pg BEQ/g wet weight for PCDD/Fs and dioxin-like PCBs.



# European Union Reference Laboratory for halogenated POPs in Feed and Food



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## 2. Participating laboratories

This proficiency test was open for participation of:

- National Reference Laboratories (NRLs) of EU member states
- National Reference Laboratories of other European countries
- Official laboratories
- Commercial laboratories

Table 1: Participating laboratories

Participating laboratories	Region	No. of participants
National Reference Laboratories	European Union Other Countries	27 2
Official Laboratories	European Union Other European Countries Africa Americas Asia Oceania	60 0 0 2 1 0
Commercial Laboratories	European Union Other European Countries Africa Americas Asia Oceania	16 1 0 2 2 0
	Total	113

### 2.1 Number of reported results

Table 2: Reported results for PCDD/F and PCB sum parameters

Reported results	WHO-PCDD/F-PCB-TEQ	WHO-PCDD/F-TEQ	WHO-PCB-TEQ	Sum of six indicator PCBs	PCDD/F-PCB-BEQ [Bioanalytical screening methods]
Fish fillet (2001-FI)	78	78	79	104	10



## 2.2 Accreditation

Table 3: Reported accreditation according to ISO/IEC 17025 by participants for PCDD/Fs and PCBs

Accreditation according to ISO/IEC 17025	PCDD/Fs, PCBs [Physico-chemical methods]	PCDD/Fs, PCBs [Bioanalytical screening methods]
yes	102	7
no	7	3

## 3. Detection methods

The following detection methods were applied:

- GC-HRMS-, GC-MS/MS-, GC-LRMS-methods for PCDD/Fs and non-ortho PCBs
- GC-HRMS-, GC-MS/MS-, GC-LRMS-, GC-ECD-methods for mono-ortho-PCBs and indicator PCBs
- Bioanalytical screening methods for PCDD/Fs and dioxin-like PCBs

Table 4: Overview of physico-chemical detection methods for PCDD/Fs and PCBs in fish fillet (2001-FI)

Detection methods	PCDD/Fs	non-ortho-PCBs	mono-ortho-PCBs	Indicator PCBs
HRMS	67	67	63	51
MS/MS	8	9	10	29
LRMS	3	3	2	10
ECD	-	-	1	8



#### 4. Homogeneity and stability of the test material

The test for sufficient homogeneity was performed according to ISO 13528:2015 [2] and the International Harmonized Protocol for the Proficiency Testing of Analytical Chemistry Laboratories [1].

Therefore, 10 portions of the test sample 2001-FI were analyzed in duplicate for PCDD/Fs and PCBs. The test for sufficient homogeneity was performed for the sum parameters WHO-PCDD/F-PCB-TEQ, WHO-PCDD/F-TEQ, WHO-PCB-TEQ, the sum of six indicator PCBs and PCDD/F, PCB and PBDE congeners. The test materials showed sufficient homogeneity for this proficiency test.

The stability check of the analytes of interest applying room temperature storage was performed according to ISO 13528:2015 [2]. The test materials showed sufficient stability for this proficiency test.

#### 5. Determination of the assigned values

Statistical evaluation of the PT results is performed by the EURL for Halogenated POPs in Feed and Food according to ISO 13528:2015 [2] and the International Harmonized Protocol for the Proficiency Testing of Analytical Chemistry Laboratories [1].

The determination of the assigned value is performed according to [1] by estimating of the assigned value as the consensus of participants' results (using only results of physico-chemical methods). The Huber robust mean is taken as assigned value after excluding extreme outliers (outside the range of  $\pm 50\%$  of the median of all reported results) and examination of the distribution of the remaining results using histogram and kernel density estimation, if necessary.

The assigned value is calculated for WHO-PCDD/F-PCB-TEQ, WHO-PCDD/F-TEQ, WHO-PCB-TEQ, the sum of six indicator PCBs and individual PCDD/F and PCB (including limits of quantification (LOQs)), if possible. Additionally the median of all values is calculated.

For individual congeners (including LOQs) assigned values are only calculated according to the above mentioned procedure, if more than 2/3 of all results are above the LOQ and less than 1/3 of all results (including LOQs) are outside the range of  $\pm 50\%$  of the median of all reported results. Levels for individual congeners are only taken for, if these levels are equal to or above the LOQ; otherwise the LOQ will be taken.

Assigned values could not be calculated for the lower bound WHO-PCDD/F-TEQ and 9 PCDD/F congeners due to the high variation of participants' results or the limited number of reported results above the LOQ.

Since there are no traceable reference values available, the assigned values in this PT were calculated on the basis of the Huber robust mean of the results of the participants.



# European Union Reference Laboratory for halogenated POPs in Feed and Food



State Institute for Chemical and Veterinary Analysis of Food, Freiburg, Germany

Therefore, the assigned values are only traceable to the results of the participants. Additionally the results of all participants reporting results and the results of participants having accreditation according ISO/IEC 17025 were compared for PCDD/F and PCB sum parameters. No significant differences between the assigned values calculated for both data sets were observed.

Table 5: Comparison of assigned values for all participants and participants with reported accreditation according to ISO/IEC 17025 for PCDD/F, PCB and PBDE sum parameters

<b>Test sample</b>	<b>Assigned value</b>		<b>Deviation</b>
	All participants	ISO/IEC 17025 accreditation	
	pg/g, ng/g wet weight	pg/g, ng/g wet weight	%
WHO-PCDD/F-PCB-TEQ ub rep	2.11	2.09	1
WHO-PCDD/F-PCB-TEQ lb rep	2.05	2.04	0
WHO-PCDD/F-TEQ ub rep	0.216	0.216	0
WHO-PCDD/F-TEQ lb rep	-	-	-
WHO-PCB-TEQ ub rep	1.85	1.86	1
WHO-PCB-TEQ lb rep	1.84	1.85	1
Sum Indicator PCBs ub rep	39.0	39.2	1
Sum Indicator PCBs lb rep	38.7	39.0	1



## 5.1 PCDD/Fs and PCBs – Sum parameters

The assigned values for the test sample 2001-FI were calculated as consensus of participants' results for the PCDD/F and PCB sum parameters.

Table 6: Assigned values for physico-chemical methods for PCDD/Fs and PCBs (rounded to three significant figures)

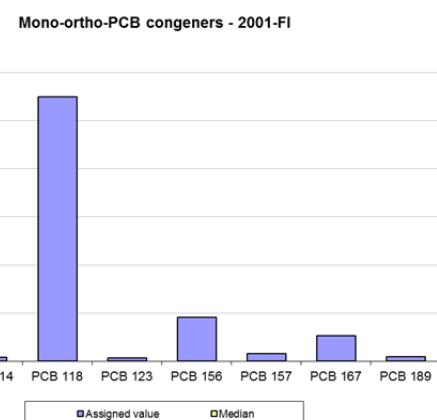
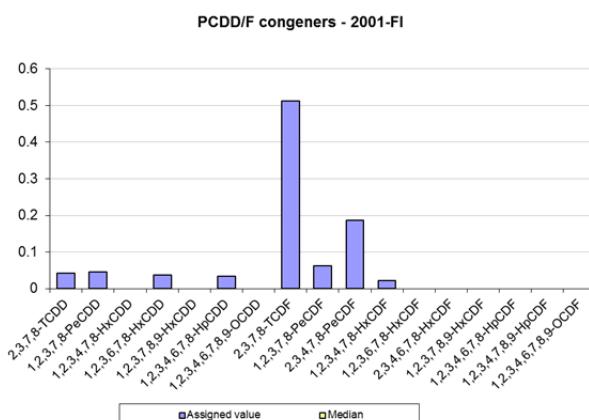
Test sample	WHO-PCDD/F-PCB-TEQ upper bound	WHO-PCDD/F-TEQ upper bound	WHO-PCB-TEQ upper bound	Sum Indicator PCBs upper bound
	pg/g wet weight	pg/g wet weight	pg/g wet weight	ng/g wet weight
Fish fillet (2001-FI)	2.11	0.216	1.85	39.0

Table 7: Assigned values for PCDD/Fs and DL-PCBs for comparison with BEQ results of bioanalytical screening methods (rounded to two significant figures)

Test sample	WHO-PCDD/F-PCB-TEQ upper bound	WHO-PCDD/F-TEQ upper bound	WHO-PCB-TEQ upper bound
	pg/g wet weight	pg/g wet weight	pg/g wet weight
Fish fillet (2001-FI)	2.1	0.22	1.8

## 5.2 PCDD/Fs and PCBs – Individual congeners

The assigned values for the test sample 2001-FI for individual congeners were calculated as consensus of participants' results taken into account criteria for calculation as described above.





# European Union Reference Laboratory for halogenated POPs in Feed and Food



State Institute for Chemical and Veterinary Analysis of Food, Freiburg, Germany

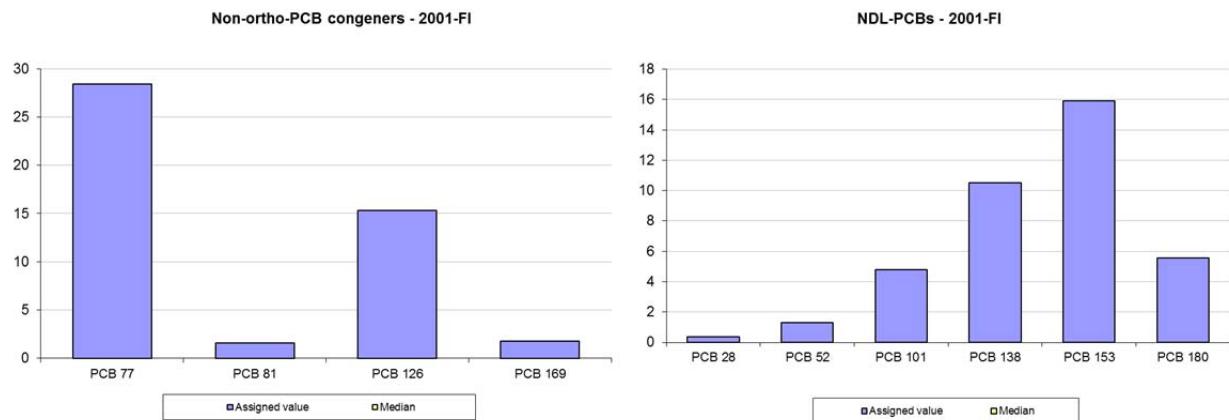


Figure 1: Assigned (blue) for PCDD/F and PCB congeners for fish fillet (2001-FI) [pg/g and ng/g wet weight]

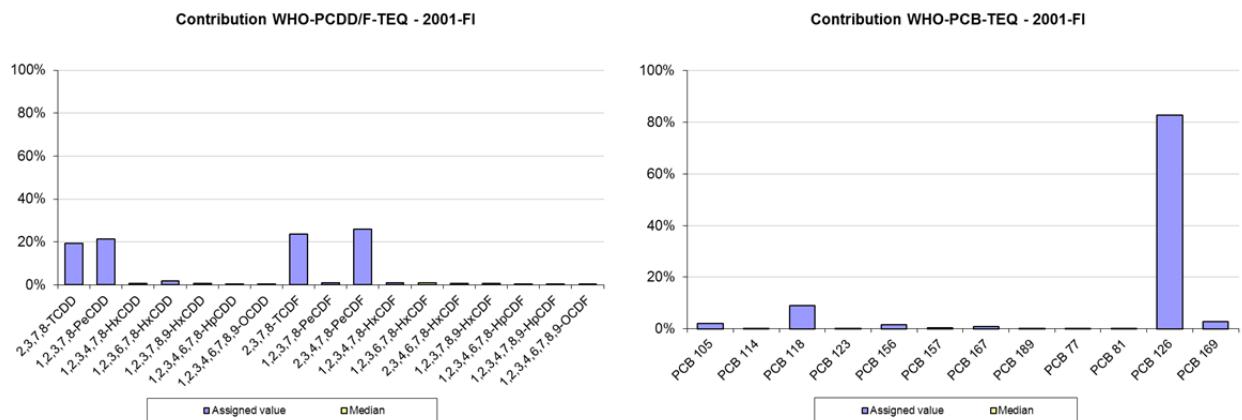


Figure 2: Contributions in % to WHO-PCDD/F-TEQ and WHO-PCB-TEQ for PCDD/F and PCB assigned and median values for fish fillet (2001-FI)



### 5.3 Comparison of assigned values with legal limits

Maximum levels for food are defined in Commission Regulation (EC) No 1881/2006 of 19 December 2006 setting maximum levels for certain contaminants in foodstuff. Action levels are defined in Commission Recommendation of 3 December 2013 on the reduction of the presence of dioxins, furans and PCBs in feed and food (2013/711/EU)

Table 8: Maximum and action levels for muscle meat of fish:

Muscle meat of fish and fishery products and products thereof	Maximum level	Action level
WHO-PCDD/F-PCB-TEQ	pg/g wet weight	6.0
WHO-PCDD/F-TEQ	pg/g wet weight	3.5
WHO-PCB-TEQ	pg/g wet weight	-
Sum of 6 Indicator PCBs	ng/g wet weight	75 <sup>(1)</sup> / 125 <sup>(2)</sup>

<sup>(1)</sup> Muscle meat of fish and fishery products and products thereof, with the exemption of wild caught fresh water fish, with the exception of diadromous fish species caught in fresh water

<sup>(2)</sup> Muscle meat of wild caught fresh water fish with the exception of diadromous fish species caught in fresh water, and products thereof

<sup>(3)</sup> Muscle meat of farmed fish and farmed fishery products

For the test sample 2001-FI the assigned values for the sum parameters WHO-PCDD/F-PCB-TEQ, WHO-PCDD/F-TEQ, WHO-PCB-TEQ and the sum indicator PCBs were below the respective maximum levels and/or action levels.

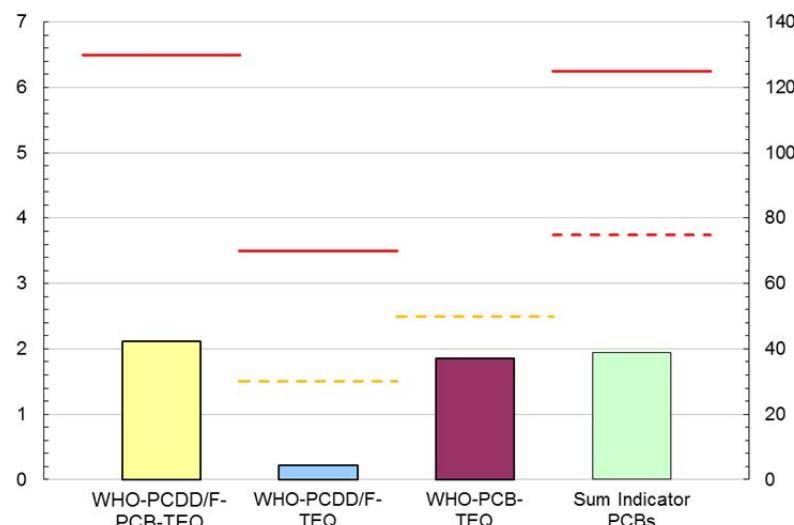


Figure 3: Comparison of assigned values for sum parameters for fish fillet (2001-FI) with maximum levels (red line) and action levels (yellow line) [pg/g and ng/g wet weight]



## 6. Evaluation of results

### 6.1 Physico-chemical methods

#### 6.1.1 Z-score calculation

Criteria for successful participation of laboratories using physico-chemical methods are based on the evaluation of the results of the sum parameters WHO-PCDD/F-TEQ, WHO-PCB-TEQ, WHO-PCDD/F-PCB-TEQ and the sum of six indicator PCBs and evaluated individual congeners. The criteria will be applicable for sum parameter concentrations in the range (about 0.5 to 4 times) of the level of interest (maximum or action level).

For evaluation of results of physico-chemical methods the z-scores are calculated according to the following formula:

$$z = (x - x_a) / \sigma_p$$

$x_a$ : assigned value

$x$ : participants result

$\sigma_p$ : fitness-for-purpose-based standard deviation for proficiency assessment

For WHO-PCDD/F-TEQ, WHO-PCB-TEQ and WHO-PCDD/F-PCB-TEQ the standard deviation for proficiency assessment  $\sigma_p$  is defined as 10 %, for the sum of six indicator PCBs (PCB 28, 52, 101, 138, 153, 180) as 15 % and for evaluated individual PCDD/F and PCB as 20 %.

Z-scores for individual congeners and diastereomers are only calculated and reported if levels for these congeners are equal to or above the LOQ. Otherwise no z-scores will be given.

Interpretation of z-scores:

$ z\text{-score}  \leq 2$	satisfactory performance
$2 <  z\text{-score}  < 3$	questionable performance (warning signal)
$ z\text{-score}  \geq 3$	unsatisfactory performance (action signal)



### 6.1.2 PCDD/Fs and PCBs - Participants' z-scores

Concentrations for WHO-PCDD/F-PCB-TEQ, WHO-PCDD/F-TEQ, WHO-PCB-TEQ and the sum of six indicator PCBs in the test sample 2001-FI are below the range (about 0.5 to 4 times) of the respective maximum levels set for muscle meat of wild caught fresh water fish. The level of the sum of six indicator PCBs is within the range of the respective maximum level for muscle meat of fish and fishery products and products thereof, with the exemption of wild caught fresh water fish. Action levels for WHO-PCDD/F-TEQ and WHO-PCB-TEQ are only set for muscle meat of farmed fish and farmed fishery products. The levels for WHO-PCB-TEQ in the PT sample are within the range of this action level.

Table 9: Distribution of participants' z-scores for sum parameters for fish fillet (2001-FI)

Percentage of participants' results	WHO-PCDD/F-PCB-TEQ	WHO-PCDD/F-TEQ	WHO-PCB-TEQ	Sum of six indicator PCBs
$ z\text{-score}  \leq 2$	78 %	45 %	77 %	82 %
$2 <  z\text{-score}  < 3$	9 %	19 %	5 %	10 %
$ z\text{-score}  \geq 3$	13 %	36 %	18 %	8 %

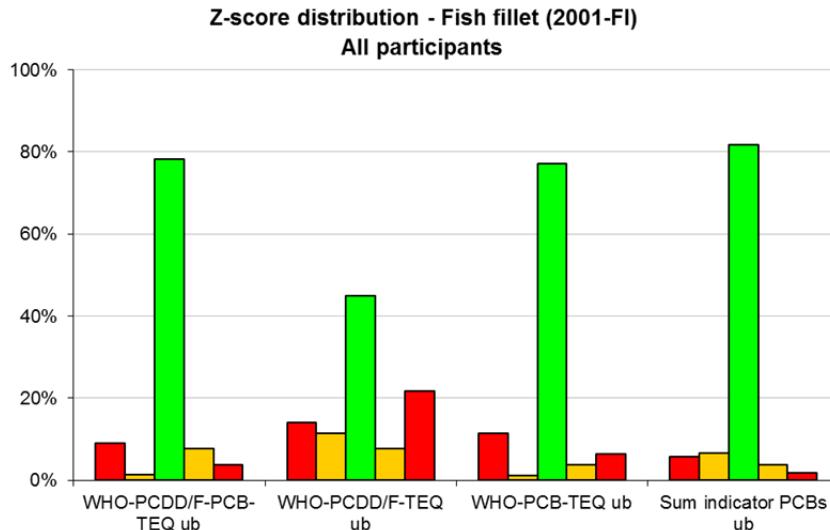


Figure 4: Distribution of participants' z-scores for sum parameters for fish fillet (2001-FI)  
[Green bars:  $-2 \leq z\text{-score} \leq 2$ , yellow bars:  $-3 < z\text{-score} < -2$ ,  $2 < z\text{-score} < 3$ , red bars:  $z\text{-score} \leq -3$ ,  $z\text{-score} \geq 3$ ]



### 6.1.3 PCDD/Fs and PCBs - Positive scoring system

The “positive scoring system” gives one assessment for each PT sample covering all relevant PCDD/F and PCB sum parameters and congeners.

The total score for the positive scoring system is calculated according to the following general principles:

- Calculation of z-scores for sum parameters and evaluated individual congeners
- Calculation of the positive scores according to the following table:

Table 10: Positive scores allocated to z-scores achieved for all parameters

Positive scoring system	z-score   ≤ 2	2 <   z-score   < 3	z-score   ≥ 3
Individual congeners	Positive score	Positive score	Positive score
Contribution to sum parameter* > 10 %	12	6	0
Contribution to sum parameter* 3 – 10 %	8	4	0
Contribution to sum parameter* < 3 %	6	3	0
Not evaluated congeners	0	0	0

\*separately for the respective sum parameters WHO-PCDD/F-TEQ, WHO-PCB-TEQ and the sum of six indicator PCBs

- Calculation of maximum achievable scores ( $| z-score | \leq 2$ ) for PCDD/F and DL-PCB and indicator PCB congeners separately:

$$Maximum Score = \sum_{i=1}^n Max.Score_{(>10\%)} + \sum_{i=1}^m Max.Score_{(3-10\%)} + \sum_{i=1}^p Max.Score_{(<3\%)}$$

- Calculation of the participants' scores for PCDD/F and DL-PCB and indicator PCB congeners separately:

$$Participant's Score = \sum_{i=1}^n Score_{(>10\%)} + \sum_{i=1}^m Score_{(3-10\%)} + \sum_{i=1}^p Score_{(<3\%)}$$

- Calculation of achieved scoring percentage for each participant:

$$Participant's Scoring Percentage = \frac{Participant's score}{Maximum score} \cdot 100$$



# European Union Reference Laboratory for halogenated POPs in Feed and Food



State Institute for Chemical and Veterinary Analysis of Food, Freiburg, Germany

- Criteria for successful participation:

Sum parameters:	≤ 1 parameter with   z-score   > 2, no parameter with   z-score   ≥ 3
PCDD/F congeners:	≥ 75 % of maximum score
DL-PCB congeners:	≥ 75 % of maximum score
Indicator PCB congeners:	≥ 75 % of maximum score
Difference between reported and calculated results for sum parameters	≤ 10 %

The assessment based on the positive scoring system is performed for each PT test sample. A laboratory participates successfully in a PT for PCDD/Fs and PCBs, if all above mentioned criteria for the reported analytes are met for each PT test sample.

Table 11: Successful participation rate according to positive scoring system for fish fillet (2001-FI)

Scoring system	Successful participation		Reason for not successful participation			
	yes	no	Only sum parameters	Sum parameters + individual congeners	Only individual congeners	Calculation of sum parameters
2001-FI	73 %	27%	35%	32%	23%	29%

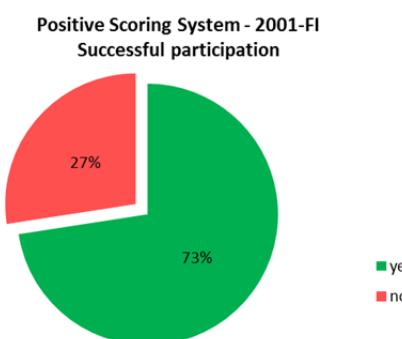


Figure 5: Successful participation rate according to the positive scoring system for fish fillet (2001-FI) [%]



#### 6.1.4 Comparison of reported and calculated sum parameters

In addition to the calculation of the sum parameters for reported individual PCDD/F and PCB congener values, the calculated sum parameters for PCDD/Fs and PCBs by the EURL are compared with the ones reported by each participant. As the reported sum parameters are decisive to compare the results with the legal limits, an incorrect calculation might lead to a wrong assessment of a sample. Therefore, in case of a significant deviation of the reported sum parameter value from the (EURL) calculated one (deviation > 10 %), the respective results will be marked as incorrect and no z-score will be given in the certificate of participation.

Table 12: Difference between reported and calculated sum parameters for PCDD/Fs and PCBs for fish fillet (2001-FI): Percentage of participants' results

Difference between reported and calculated sum parameters	WHO-PCDD/F-PCB-TEQ	WHO-PCDD/F-TEQ	WHO-PCB-TEQ	Sum of six indicator PCBs
Deviation ≤ 10 %	98 %	96 %	99 %	99 %
Deviation > 10 %	2 %	4 %	1 %	1 %



### 6.1.5 Difference between upper and lower bound calculation

According to Commission Regulation (EU) No 2017/644 the difference between upper bound level and lower bound level shall not exceed 20 % for confirmation of exceedance of maximum level or in case of need of action levels for PCDD/Fs and DL-PCBs. For indicator PCBs the difference between upper bound and lower bound levels for the sum of six indicator PCBs shall be  $\leq 20$  % at the level of interest.

For the test sample 2001-FI the assigned values for all sum parameters are in the range or below the respective maximum or action levels.

Table 13: Difference between upper and lower bound calculation for fish fillet (2001-FI): Percentage of participants' results

Percentage of participants' results	WHO-PCDD/F-PCB-TEQ	WHO-PCDD/F-TEQ	WHO-PCB-TEQ	Sum of six indicator PCBs
0 – 10 %*	97 %	95 %	97 %	94 %
10 – 20 %*	0 %	0 %	1 %	6 %
20 – 50 %*	0 %	0 %	1 %	0 %
> 50 %*	3 %	3 %	3 %	0 %

\* Difference between upper and lower bound calculation

Difference between upper bound and lower bound calculation - 2001-FI

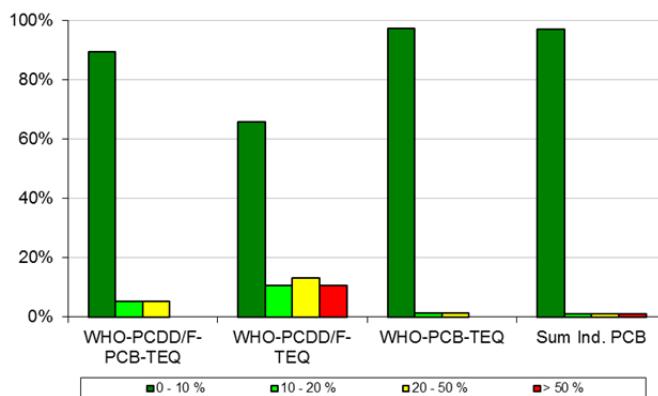


Figure 6: Difference between upper and lower bound calculation for fish fillet (2001-FI): Percentage of participants' results (Dark green bars: 0 – 10 %, light green bars: 10 – 20 %, yellow bars: 20 – 50 %, red bars: > 50 %)



### 6.1.6 Assessment of analytical results and measurement uncertainty

In addition, participants were asked to report the measurement uncertainty applied for checking of compliance for WHO-PCDD/F-PCB-TEQ, WHO-PCDD/F-TEQ, WHO-PCB-TEQ and the sum of six indicator PCBs and the assessment if the analytical results for the sample exceed the respective maximum and action levels beyond reasonable doubt taking into account the measurement uncertainty.

Table 14: Reported relative expanded measurement uncertainty for sum parameters for fish fillet (2001-FI); outlier removed (e.g. reporting of absolute values)

Reported measurement uncertainty	WHO-PCDD/F-PCB-TEQ	WHO-PCDD/F-TEQ	WHO-PCB-TEQ	Sum of six indicator PCBs
Minimum	7 %	6 %	6 %	2 %
Mean	24 %	23 %	24 %	24 %
Median	20 %	21 %	20 %	23 %
Maximum	50 %	50 %	50 %	53 %

Table 15: Percentage of participants reporting exceedance of legal limits fish fillet (2001-FI)

Participants reporting exceedance of legal limit	WHO-PCDD/F-PCB-TEQ	WHO-PCDD/F-TEQ	WHO-PCB-TEQ	Sum of six indicator PCBs
Maximum level	1 %	0 %	-	0 %
Action level	-	0 %	3 %	-

## 6.2 Bioanalytical screening methods

According to Commission Regulation (EU) No 2017/644, “a screening method in principle classifies a sample as compliant or suspected to be non-compliant. For this, the calculated BEQ level is compared to the cut-off value [...]. Samples below the cut-off value are declared compliant, samples equal or above the cut-off value as suspected to be non-compliant, requiring analysis by a confirmatory method”.

Therefore, the main criterion for evaluation of results from bioanalytical screening methods is their ability to reliably identify compliant samples and samples suspected to be non-compliant with established legal limits.

For further evaluation of the performance of bioanalytical screening methods, bioassay-scores are applied: The reported BEQ-values derived from bioanalytical screening methods are compared with the WHO-TEQ assigned values calculated on basis of the results of physical-chemical methods for the concentration range of 0.5 to 2 times the level of interest. Because bioanalytical screening methods focus mainly on distinguishing be-



tween compliant and potentially non-compliant samples, a direct comparison of bioassay-scores and z-scores is not possible. However, bioassay scores may serve as a tool to assess method performance within the scope of external quality control measures of the respective laboratory.

Bioassay-scores are calculated according to the following formula:

$$\text{bioassay-score} = (x - x_a) / \sigma_{\text{bioassay}}$$

$x_a$ : assigned value (physical-chemical methods)

$x$ : participants result (BEQ from bioanalytical screening method)

$\sigma_{\text{bioassay}}$ : bioassay target deviation

For PCDD/F-BEQ, PCB-BEQ and PCDD/F-PCB-BEQ the bioassay target deviation  $\sigma_{\text{bioassay}}$  is defined as 20 %.

### 6.2.1 Assessment of analytical results

As a consequence of the comparison of assigned values with legal limits, the assessment of the analytical results using bioanalytical screening methods should read “suspected to be non-compliant with the maximum level for WHO-PCDD/F-PCB-TEQ, WHO-PCDD/F-TEQ and the action level for WHO-PCB-TEQ” for the test sample 2001-FI.

Table 16: Participants' assessment of analytical results using bioanalytical screening methods for fish fillet (2001-FI)

Laboratories' assessment of analytical results	WHO-PCDD/F-PCB-TEQ Maximum level	WHO-PCDD/F-TEQ Maximum level	WHO-PCDD/F-TEQ Action level	WHO-PCB-TEQ Action level
Suspected to be non-compliant	1	2	2	2
Compliant	7	5	4	4



### 6.2.2 Participants' bioassay-scores

Concentrations for WHO-PCDD/F-PCB-TEQ, WHO-PCDD/F-TEQ and WHO-PCB-TEQ in the test sample 2001-FI below the range (about 0.5 to 2 times) of the respective maximum and action levels. Only the range of the action level for muscle meat of farmed fish and farmed fishery products is covered.

Table 17: Distribution of participants' bioassay-scores for BEQ parameters for fish fillet (2001-FI)

Percentage of participants' results	PCDD/F-PCB-BEQ	PCDD/F-BEQ	PCB-BEQ
bioassay-score   ≤ 2	5	0	2
2 <   bioassay-score   < 3	1	1	0
bioassay-score   ≥ 3	3	3	2

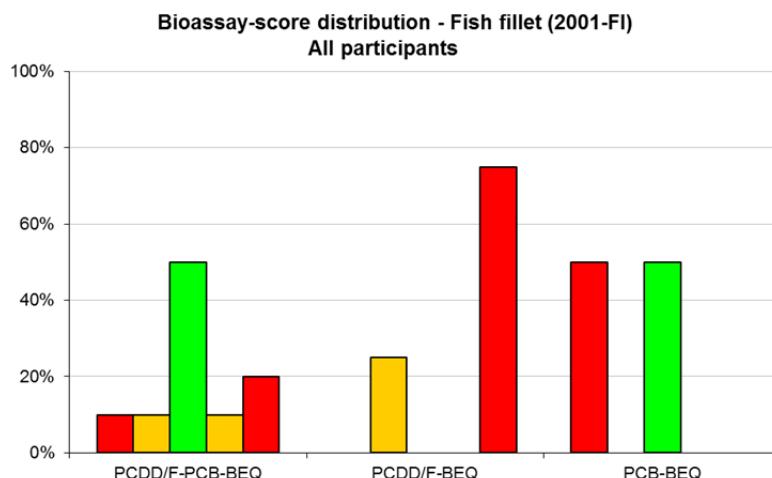


Figure 7: Distribution of participants' bioassay-scores for BEQ parameters for fish fillet (2001-FI) (Green bars:  $-2 \leq \text{bioassay-score} \leq 2$ , yellow bars:  $-3 < \text{bioassay-score} < -2, 2 < \text{bioassay-score} < 3$ , red bars:  $\text{bioassay-score} \leq -3, \text{bioassay-score} \geq 3$ )



## 7. Participants' feedback

A questionnaire for feedback from participants of this EURL proficiency test was available as online survey between 23 July 2020 and 31 August 2020. The survey was anonymous, but participants could also give their laboratory name. The identity of the laboratories is kept confidential. The survey included seven questions related to different topics (participants' information, organization of the proficiency test, PT test samples and evaluation of results and summary of data) and a possibility to include comments and further suggestions. In total, 20 laboratories (18 % of all participants) participated in this survey. A summary of the results is also given in annex 8.

### 7.1 Overview of questions and answers of participants

#### Participants' information:

National Reference Laboratory (NRL)	Official Laboratory (OFL)	Commercial laboratory	Other
63 %	5 %	26 %	11 %

#### Organization of proficiency test:

	Fully	Largely	Partly	Not at all	No opinion
Satisfied with organization of PT	68 %	32 %	-	-	-
Meeting of expectations	74 %	21 %	5 %	-	-
Information understandable	68 %	32 %	-	-	-
Time frame acceptable	74 %	26 %	-	-	-

#### PT test samples:

	Fully	Largely	Partly	Not at all	No opinion
Selection of matrix and level of contamination adequate	47 %	37 %	16 %	-	-

#### Evaluation of results and summary of data:

	Fully	Largely	Partly	Not at all	No opinion
Evaluation of results and report clear and comprehensible	68 %	21 %	11 %	-	-



## 7.2 Comments and suggestions

Comments referred to the too low concentrations for PCDD/Fs and the too complex tables for the overview of the results. Additionally also the long time between preliminary and final report was mentioned.

## 8. Quality control

The Deutsche Akkreditierungsstelle GmbH attests that the provider of proficiency testing Chemisches und Veterinäruntersuchungsamt Freiburg, EU-Reference Laboratory (EURL) for halogenated persistent organic pollutants (POPs) in Feed and Food is competent under the terms of DIN EN ISO/IEC 17043:2010 to carry out proficiency testing in the testing field of determination of halogenated persistent organic pollutants (POPs) in food and feed (Accreditation number: D-EP-18625-01-00).

## 9. Summary of participants' results

An overview of the PCDD/F and PCB results for the PT test sample fish fillet (2001-FI) and the evaluation of the results are given in the following annexes 1 - 8. Laboratories are coded according to the laboratory codes sent after registration.

## 10. References

- [1] M. Thompson, S.L.R. Ellison, R. Wood: The International Harmonized Protocol For The Proficiency Testing Of Analytical Chemistry Laboratories, Pure Appl. Chem., Vol. 78, No. 1, pp. 145-196, 2006.
- [2] ISO 13528:2015, Statistical methods for use in proficiency testing by interlaboratory comparisons, International Organization for Standardization
- [3] M. van den Berg et al., The 2005 World Health Organization Re-evaluation of Human and Mammalian Toxic Equivalency Factors for Dioxins and Dioxin-like Compounds. Toxicological Sciences 93(2), 223-241 (2006)



## 11. Annex

(Please double click on the pdf-icons to open the annexes.)

<b>Fish fillet (2001-FI)</b>	
1	Assigned values – PCDD/F, PCB
2	Participants' results – Tables – PCDD/F, PCB
3	Participants' z-scores and bioassay-scores – Tables - PCDD/F, PCB
4	Participants' z-scores – Charts – PCDD/Fs, PCB
5	Scoring system – PCDD/F, PCB
6	Homogeneity and stability test – PCDD/F, PCB
7	Participants' methods – PCDD/F, PCB

<b>Questionnaire for feedback from participants</b>	
8	Summary of feedback

EURL for halogenated Persistent Organic Pollutants (POPs) in Feed and Food  
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